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Antimicrobial Activity and Chemical Composition of Pistachia Atlantica Gum Sub Sp. Kurdica. Essential Oil

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ABSTRACT

Background: Staphylococcus aureus, Salmonella enterica, Escherichia Coli (E. Coli) and Listeria monocytogenes are considered as important foodborne pathogens. Pistachia atlantica sub sp. Kurdica, called wild pistachio, has been known as an antimicrobial compound. The aim of this study was to determine the antimicrobial activity and chemical composition of this essential oil (EO) on some of foodborne pathogens. Methods: The EO of Pistachia atlantica was obtained by hydro-distillation and analyzed by GC-MASS. The antibacterial effects of Pistachia atlantica were evaluated at two concentrations of 10 and 15 μL against Staphylococcus aureus, E. Coli, Salmonella enterica, and Listeria monocytogenes using disk diffusion method. The analysis was done by SPSS. Results: In the current study, α-pinene (92.5%) and β-pinene (1.62%) were the main components of Pistachia atlantica EO. The EO was most effective on Salmonella enterica, whereas, its effect on Listeria monocytogenes was the weakest. The results showed a significant difference in reducing Salmonella enterica in comparison to others (P < 0.05). Conclusion: The EO has inhibitory effects on the studied bacteria. Therefore, this EO can be used as a natural preservative to extend the shelf life of foods.

Keywords: Pistachia atlantica; antimicrobial; Listeria monocytogenes; Staphylococcus aureus; Escherichia Coli; Listeria monocytogenes

Introduction

Food can be contaminated by pathogenic bacteria including Salmonella enterica, Staphylococcus aureus, Escherichia Coli (E. Coli),

and *Listeria monocytogenes*, which can cause several problems in gastro intestinal tract. During the last decades, extensive use of antimicrobial

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drugs has led to bacterial resistance in human beings (Moshafi et al., 2004). Essential oils (EO) are aromatic oils extracted from plants by different methods (Burt, 2004). Essential oils have been recognized as natural and harmless antimicrobial agents for the environment and Generally Recognized As Safe (GRAS) in food application (Nawel et al., 2013). The antibacterial effects of EOs depend on the type of plant, meteorological conditions, place of growth, as well as drying and processing methods. The use of oil plants in Iran as an antioxidant or antibiotic was traced back to ancient times (Akrami et al., 2015). Pistachio atlantica subsp. Kurdica, called Baneh, is one of the wild pistachio species (grows in Zagros forests), which belongs to the Anacardiacea family (Karimi et al., 2009). Three subspecies of cabulica, kurdica, and mutica grow in Iran. Over 1,200,000 hectares of western, central, and eastern parts of Iran are covered by kurdica and mutica subspecies (Farhoosh et al., 2008). Pistacia atlantica tree grows in the western (mainly), central, and eastern part of Iran. The local people use this tree to produce ingredients in some food, jams, and gum (Hatamnia et al., 2014). The gum obtained from this tree consists of oleoresin. This essential oil contains α-Thujene, α-Pinene, Camphorene, Sabinene, β-Pinene, Δ3-Carene Limonene, as well as alphatropineol, armanderen, and caprylic acid (Sharifi and Hazell, 2011). The results of previous studies show that the presence of resin in Pistachio has an inhibitory effect on the growth of clostridium botulinum, Staphylococcus aureus, E. Coli, and Streptococcus (Aksoy et al., 2006, Daifas et al., 2004). In another study, the antimicrobial activity of Pistachio atlantica Subsp., i.e., kurdica species were found to have positive results on helicobacter pylori (Sharifi and Hazell, 2011). The aim of the current study was to investigate the inhibition effect of Pistachio atlantica Subsp. Kurdica gum essential oil on staphylococcus aureus, Salmonella, E. Coli, and Listeria monocytogenes.

Materials and Methods

Preparation of the essential oil of Pistachia atlantica gum Subsp. Kurdica: Pistachia atlantica

sub sp. *Kurdica* gum was collected from Kordestan province in Iran. The herbarium approval of the plant was confirmed by the Institute of Medicinal Plants, Medical University of Tehran, Iran. The gum was placed inside the Clevenger along with distilled water and the extraction was carried out by steam distillation method. The ratio of the achieved essence to gum weight was also calculated. The oil was kept at low temperature for further analysis (Mohajeri *et al.*, 2018, Rahimi *et al.*, 2019).

GC-MASS analysis of essential oil: The components of gum EO were isolated by Gas Chromatography/Mass Spectrometry (GC model 9-A, Shimadzu, Japan, and GC/MS model Varian 3400, USA) using a DB-5 column with a length of 30 m and an inner diameter of 0.25 mm. Later, these components were identified by comparing their mass spectrum with the range combinations found in the computer database and valid compositions. The oven temperature was increased from 60 to 250 °C with an increment rate of 5 °C per minute. At the end, it was kept for 10 minutes at 250 °C. Helium carrier gas with a flow rate of 1.1 mL/min and ionization energy of 70 electron volts was used. The separation ratio was 1 to 50 and the injection volume was 100 µ (Rahimi et al., 2019, Sharifi and Hazell, 2011).

Determination of antimicrobial properties of Pistacia atlantica gum EO by disk diffusion method: In this study, four standard bacterial strains were used including Listeria monocytogenes (PTCC 1298), Escherichia coli (ATCC 2592). Staphylococcus aureus (ATCC 25923), Salmonella enterica (PTCC 14028). At first, a microbial suspension equivalent to 0.5 McFarland turbidity standards (approximate density of 1.5×10^8 CFU/mL) was prepared from each strain. Then, 100 uL of each bacterial suspension with a density of 10⁶ CFU/mL was cultured by spreading on the surface of the Mueller-Hinton agar medium. A 5mm diameter sterile paper disk was placed on the medium and 15 µL of pure EO was injected into the disk. Plates were incubated for 24 hours at 37° C. Then, the diameter of the inhibition zone was

measured. All treatments were carried out in triplicate (Akrami *et al.*, 2015).

Data analysis: The experimental data were

collected and analyzed using one-way ANOVA by SPSS. In all calculations, P-value of less than 0.05 was considered statistically significant.

Table1. Chemical composition of P. atlantica subsp. Kurdica gum essential oil

Type of chemical composition	Percentage	Retention Time (%)
Tricyclene	0.15	11.15
α-pinene	92.08	11.78
Camphene	0.67	12.61
Sabinene	0.36	13.79
ß-pinene	1.61	14.06
Myrcene	0.27	14.62
δ-2-carene	0.35	15.63
ρ-cymene	0.26	16.61
Limonene	0.51	16.77
1,8-cineole	0.17	16.95
Terpinolene	0.11	16.69
Verbenone	0.20	26.19
Isiborny1 acetate	0.30	29.53
Total Identified	97.02	

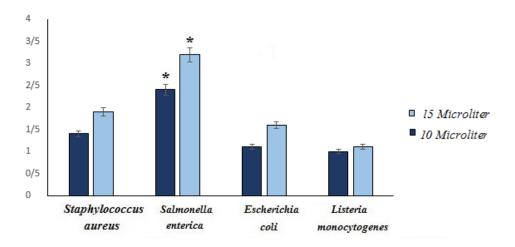


Figure 1. Growth inhibition of *Staphylococcus aureus*, *Salmonella enterica*, *Escherichia coli*, and *Listeria monocytogenes* at a concentration of 10⁶ CFU/mL by the *Pistacia atlantica* gum EO after 24 hours of exposure

Results

Chemical composition of *Pistacia atlantica* gum EO

The oil yield of the *Pistacia atlantica* gum EO was evaluated as 10% (w/w). About 97.02% of the components of gum EO were identified (approximately 13 compounds); of which α -pinene (92.08%) and β -pinene (1.61%) were predominant

(**Table 1**). Given that α -pinene was the main ingredient of *Pistacia atlantica* gum EO, the antimicrobial effects may be due to the presence of this component.

Antibacterial activity in disk diffusion method

The growth rates of Staphylococcus aureus, Salmonella enterica, Escherichia coli, and Listeria

^{*}Significant at the level of P < 0.05

monocytogenes at concentration of 10^6 CFU/mL after 24 hours of exposure with *Pistacia atlantica* gum EO are shown in **Figure 1**. The maximum inhibition zone diameter was related to *S. enterica*, while the minimum value was attributed to *L. monocytogenes*. The results showed a significant difference in reduction of *Salmonella enterica* in comparison to others (P < 0.05). This result indicated that by increasing the concentration, the diameter of inhibition zone increased significantly (P < 0.05).

Discussion

Nevertheless, making definite conclusion in this regard is difficult, since other compounds such as β-pinene, sabinene, and camphene exist in Pistacia atlantica gum EO, which may have contributed to the inhibitory effect (Delazar et al., 2003, Mecherara-Idjeri et al., 2008). Minaiyan et al. reported 38 compounds in the Pistacia atlantica tree's EO, among which α -pinene (41.23%), β pinene (6.85), and trans-verbenol (5.39%) had the highest concentrations (Minaiyan et al., 2015). In a similar study on the *Pistacia atlantica* composition in Marven region, Iran, α-pinene comprised about 70% of the EO (Delazar et al., 2003). This difference in amount may be due to the different species or geographic regions (Benhammou et al., 2008, Ghalem and Mohamed, 2009).

The agar diffusion test showed that Salmonella enterica had the highest sensitivity to the EO, while Listeria monocytogenes had the least sensitivity. So, it can be concluded that the Pistacia atlantica gum EO had an inhibitory effect on both gram positive and gram negative bacteria. In a similar study, the antimicrobial effect of Pistacia lentiscus leaf EO against gramnegative bacteria (such as pseudomonas and salmonella) and gram-positive bacteria (such Staphylococcus aureus, **Bacillus** and Enterococcus) was investigated by disk diffusion method. Salmonella and Enterobacter had a higher sensitivity to the EO (Derwich et al., 2010). The results of our study are in agreement with those reported by Gholem and Mohamed research. Their findings indicated that α-pinene had an antibacterial agent in the Pistacia atlantica's gum (Ghalem and Mohamed, 2009). The effect of EO on different bacteria may vary according to the laboratory conditions, the gum harvesting season, the gum's freshness or being processed, as well as the method equipment and of the extraction. Benhammou et al. studied the effect of Pistacia atlantica's gum on eight bacteria (E coli, Klebsiella pneumoneae, Pseudomonas aeruginosa, Salmonella typhi, Enterobacter cloacea, Proteus mirabilis, Listeria monocytogenes, and Staphylococcus aureus) using disk diffusion method. They showed that S. typhi and S. aureus had the most sensitivity, while K. pneumoneae and L. monocytogenes were the most resistant bacteria (Benhammou et al., 2008). A study on the effect of *Pistacia atlantica*'s gum EO on Streptococcus mutans by disc diffusion method showed that by increasing the concentration from 20 to 50 mg/mL, the inhibition zone diameter increased (Delazar et al., 2003).

Conclusion

The results of this study indicated that *Pistachia* atlantica subsp. kurdica had inhibitory effects on Staphylococcus aureus, Salmonella enterica, Escherichia coli, and Listeria monocytogenes. Thus, by studying the organoleptic effects of *Pistachia atlantica*, this compound can be used as a natural preservative for increasing the shelf life of food products.

Authors' contribution

Akrami, Hekmati moghaddam, Jebali, Sadeghizadeh Yazdi, and Khalili conceived and designed the experiments. Ellahi and Rastiani carried out the experiment. All authors contributed to the final version of the manuscript.

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Conflict of interests

The authors declare no conflict of interests regarding the publication of this paper.

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